

AD-A221 522

MENTATION PAGE

Form Approved OME No. 0704-0188

N OC 20113

2 REPORT DATE I ASSESSY USE CHLY (Laure blank) April 14, 1990 3. REPORT TYPE AND DATES COVERED

Final TEchnical Report 8/15/86-2/14/90

A TITLE AND SUPPLY

Biosynthesis, Physiological Disposition, and Biochemical

A RIMOUNG MILLERS

Effects of Nephrotoxic Glutathione and Cysteine S-Conjugates AFOSR-86-0302

M. W. Anders, D.V.M., Ph.D.

Principal Investigator

2312/05

61102F

TO PREPARE CHEATELY TION NAMES) AND ADDRESSED

University of Rochester Department of Pharmacology

601 Elmwood Avenue Rochester, NY 14642

6 ORGANIZATION

APOGR-TR- 190 - 0 4 9 4

A MANAGEME / MONITORINE AGENCY NAMES) AND ACCRESSES

United States Air Force Air Force Office of Scientific Research Building 410, Bolling AFB, DC 20332

TE SUBSCIENCE TARY NOTES

No limitations

11. ASSTRACT (Maximum 200 worth)

These studies established that the biosynthesis of S-(pentachlorobutadienyl)glutathione (PCBG) is catalyzed preferentially by hepatic microsomal glutathione S-transferases. PCBG is further metabolized to the diconjugate 1,4-bis(glutathion-S-y1)-1,2,3,4tetrachlorobuta-1,3-diene by hepatic cytosolic transferases. Studies on the synthesis of PCBG in the isolated, perfused rat liver showed that PCBG is eliminated in the bile at toxicologically relevant doses. The cysteine analog of PCBG S-(pentachlorobutadienyl)-L-cysteine (PCBC) is a potent nephrotoxin that damages mitochondria. PCBC, which is activated by renal mitochondrial cysteine conjugate \(\begin{align*} \text{-lyase, inhibits mitochondrial} \) protein, DNA, and RNA synthesis and destroys mitochondrial DNA, although the role of the effects in the observed mutagenicity of PCBC is unclear. Finally, preliminary studies on the intestinal absorption of PCBG indicate that the intact glutathione S-conjugate is absorbed in vivo and is cultured CaCo cells.

TE SUBJECT TERMS

Glutathione S-conjugates, cysteine S-conjugates, nephrotoxicity, hexachlorobūtadiene, S-(pentachlorobutadienyl)glutathione, S-(pentachlorobutadienyl)-L-cysteine

unclassified

18. SECURITY CLASSIFICATION

19. SECURITY CLASSIFICATION OF ARSTRACT

126. LIMITATION OF ABSTRACT

IL MUMBEL OF PAGES

TE PRICE COST

unclassified

STEED AT THE BEACH

unclassified

Standard Form 298 (Rev. 2-89)

GENERAL INSTRUCTIONS FOR COMPLETING SF 298

The Report Documentation Page (RDP) is used in announcing and cataloging reports. It is important that this information be consistent with the rest of the report, particularly the cover and title page. Instructions for filling in each block of the form follow. It is important to stay within the lines to meet optical scanning requirements.

- Block 1. Agency Use Only (Leave Blank)
- Block 2. Report Date. Full publication date including day, month, and year, if available (e.g. 1 Jan 88). Must cite at least the year.
- Block 3. Type of Report and Dates Covered. State whether report is interim, final, etc. If applicable, enter inclusive report dates (e.g. 10 Jun 87 - 30 Jun 88).
- Block 4. <u>Title and Subtitle</u>. A title is taken from the part of the report that provides the most meaningful and complete information. When a report is prepared in more than one volume, repeat the primary title, add volume number, and include subtitle for the specific volume. On classified documents enter the title classification in parentheses.
- Block 5. <u>Funding Numbers</u>. To include contract and grant numbers; may include program element number(s), project number(s), task number(s), and work unit number(s). Use the following labels:

C - Contract PR - Project
G - Grant TA - Task
PE - Program WU - Work Unit
Element Accession No.

Block 6. Author(s). Name(s) of person(s) responsible for writing the report, performing the research, or credited with the content of the report. If editor or compiler, this should follow the name(s).

- Block 7. Performing Organization Name(s) and Address(es). Self-explanatory.
- Block 8. <u>Performing Organization Report Number</u>. Enter the unique alphanumeric report number(s) assigned by the organization performing the report.
- Block 9. Sponsoring/Monitoring Agency Names(s) and Address(es). Self-explanatory.
- Block 10. Sponsoring/Monitoring Agency. Report Number. (If known)

Block 11. Supplementary Notes. Enter information not included elsewhere such as: Prepared in cooperation with...; Trans. of ..., To be published in When a report is revised, include a statement whether the new report supersedes or supplements the older report.

Block 12a. <u>Distribution/Availablity Statement.</u>
Denote public availability or limitation. Cite any availability to the public. Enter additional limitations or special markings in all capitals (e.g. NOFORN, REL, ITAR)

DOD - See DoDD 5230.24, "Distribution Statements on Technical Documents."

DOE - See authorities

NASA - See Handbook NHB 2200.2.

NTIS - Leave blank.

Block 12b. Distribution Code.

DOD - DOD - Leave blank
 DOE - Enter DOE distribution categories from the Standard Distribution for Unclassified Scientific and Technical

Unclassified Scientific and Technical Reports

NASA - NASA - Leave blank NTIS - NTIS - Leave blank.

Block 13. Abstract. Include a brief (Maximum 200 words) factual summary of the most significant information contained in the report.

Block 14. Subject Terms. Keywords or phrases identifying major subjects in the report.

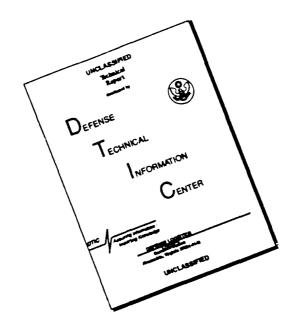
Block 15. <u>Number of Pages</u>. Enter the total number of pages.

Block 16. <u>Price Code</u>, Enter appropriate price code (NTIS only).

Blocks 17. - 19. Security Classifications.
Self-explanatory. Enter U.S. Security
Classification in accordance with U.S. Security
Regulations (i.e., UNCLASSIFIED). If form
contains classified information, stamp
classification on the top and bottom of the page.

Block 20. Limitation of Abstract. This block must be completed to assign a limitation to the abstract. Enter either UL (unlimited) or SAR (same as report). An entry in this block is necessary if the abstract is to be limited. If blank, the abstract is assumed to be unlimited.

DISCLAIMER NOTICE



THIS DOCUMENT IS BEST QUALITY AVAILABLE. THE COPY FURNISHED TO DTIC CONTAINED A SIGNIFICANT NUMBER OF PAGES WHICH DO NOT REPRODUCE LEGIBLY.

FINAL TECHNICAL REPORT

Project Title: Biosynthesis, Physiological Disposition, and Biochemical

Effects of Nephrotoxic Glutathione and Cysteine S-

Conjugates

Project Number: AFOSR-86-0302

Principal Investigator: M. W. Anders

Period Covered: 8/15/86 to 2/14/90

Biosynthesis of S-(pentachlorobutadienyl)glutathione (PCBG): The metabolism of hexachlorobutadiene (HCBD) to PCBG was studied with rat hepatic microsomal and cytosolic fractions. The major product formed was PCBG, which was fully characterized by ¹H and ¹³C NMR and by fast-atom bombardment mass spectrometry; the microsomal glutathione S-transferase was a more efficient catalyst of the formation of PCBG than were the cytosolic transferases. PCBG was metabolized to the diconjugate 1,4-bis(glutathion-S-yl)-1,2,3,4-tetrachlorobuta-1,3-diene by the cytosolic transferases. Although PCBG is nephrotoxic, insufficient diconjugate is available for toxicity studies. This work has been published (Dekant et al., 1988).

The biosynthesis of PCBG was also studied in the isolated, perfused rat liver. HCBD was metabolized to PCBG, which was almost exclusively eliminated in the bile at nonhepatotoxic doses of HCBD; when hepatotoxic doses of HCBD were studied, PCBG was eliminated in both the bile and in the caval effluent. Depletion of hepatic glutathione concentrations decreased PCBG biosynthesis in the perfused liver. This work has been submitted for public wor (Gietl and Anders, 1990).

Attempts to study the <u>in vivo</u> biosynthesis of [35S]PCBG from HCBD by labeling the hepatic glutathione pool by giving [35S]methionine failed, because the degree of labeling that could be attained was not sufficient. This strategy, which would have broad applicability, merits further investigation.

Alternative Routes of Reactive Metabolites of PCBG: In order to investigate the bioactivation mechanism of S-(pentachlorobutadienyl)-L-cysteine (PCBC), we sought alternative strategies to generate reactive intermediates of PCBC. Hence we prepared benzyl pentachlorobutadienyl sulfide, the hypothesis being that cytochromes P-450 would metabolize the sulfide to the corresponding hemimercaptal, which would eliminate the same reactive intermediate that is formed by the action of cysteine conjugate β-lyase on PCBC. The expectation was correct: benzyl pentachlorobutadienyl sulfide was cytotoxic in isolated rat hepatocytes and was metabolized to benzaldehyde by purified,

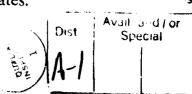
reconstituted cytochrome P-450_{PBB}. The analog <u>tert</u>-butyl pentachlorobutadienyl sulfide, which cannot form a hemimercaptal, was not, as expected, cytotoxic. This work has been published (Veltman et al., 1989). Benzyl pentachlorobutadienyl sulfide is mutagenic in the Ames test and requires activation by cytochromes P-450; this work has been published (Vamvakas et al., 1989).

In other experiments, not yet submitted for publication, we prepared the proreactive intermediate 2-nitrophenyl pentachlorobutadienyl disulfide. Reduction of the disulfide affords pentachlorobutadienylthiol, the putative reactive intermediate formed by the β-lyase-catalyzed metabolism of PCBC. We are presently attempting to characterize fully the products formed. Such stable, synthetically accessible precursors of reactive intermediates will find utility in exploring the properties of biological reactive intermediates.

Mitochondrial Toxicity of PCBC: Previous studies showed that mitochondrial are the primary targets for nephrotoxic cysteine conjugates. Hence the effect of PCBC on renal mitochondrial protein, RNA, and DNA synthesis was studied. PCBC inhibited mitochondrial protein synthesis, and this effect was blocked by the β-lyase inhibitor aminooxyacetic acid. Similarly, PCBC inhibited mitochondrial RNA and DNA synthesis, and this effect was also blocked by aminooxyacetic acid. PCBC also damages the mitochondrial genome. In mitochondria incubated with PCBC, 60% of the supercoiled, high-molecular weight mtDNA was degraded to the relaxed circular form and to small fragments. Because this effect was shared with nonmutagenic cysteine conjugates, it probably does not contribute to the nephrocarcinogenicity of HCBD. This work has been published (Banki and Anders, 1989).

Intestinal Absorption of PCBG: In work completed at the end of the period of support, we found that PCBG infused into the bile duct of rats is absorbed largely intact and can be detected in blood; PCBC was also detected in blood, indicating that some hydrolysis does take place. In studies with cultured intestinal CaCo cells, PCBG is actively transported as the intact glutathione S-conjugate. We hope to complete these studies with other funds.

<u>Summary</u>: These studies have provided new information about the biosynthesis of the glutathione conjugate PCBG and about the mitochondrial toxicity of the nephrotoxic cysteine analog PCBC. Preliminary studies indicate that PCBG is absorbed intact from the intestine. A particularly significant accomplishment was the development of proreactive intermediates of cysteine conjugates; these compounds are expected to find utility in investigating cytotoxic and mutagenic glutathione conjugates.



Publications:

Abstracts:

- Dekant, W., and Anders, M. W. (1988) Enzymatic conjugation of hexachloro-1,3-butadiene with glutathione: Formation of 1-(glutathion-\(\Sec\bar{S}\)-1,2,3,4,4-pentachloro-1,3-butadiene and 1,4-bis(glutathion-\(\Sec\bar{S}\)-1,2,3,4-tetrachloro-1,3-butadiene. FASEB J. 2, A1794.
- Dekant, W., Urban, G., and Anders, M. W. (1990) Reactive intermediates formed by the cysteine conjugate β-lyase catalyzed cleavage of colorovinyl and fluoroalkyl cysteine S-conjugates. Adv. Exp. Biol. Med., in press.
- Gietl, Y. S., and Anders, M. W. (1990) Formation and excretion of the glutathione <u>S</u>-conjugate of hexachlorobutadiene in the perfused rat liver. <u>Toxicologist</u> 10, 199.

Peer-reviewed publications:

- Banki, K., and Anders, M. W. (1989) Inhibition of rat kidney mitochondrial DNA, RNA, and protein synthesis by halogenated cysteine S-conjugates. Carcinogenesis 10, 767-772.
- Dekant, W., Vamvakas, S., Henschler, D., and Anders, M. W. (1988) Enzymatic conjugation of hexachloro-1,3-butadiene with glutathione: Formation of 1-(glutathion-\(\S_{\text{-yl}}\))-1,2,3,4,4- pentachlorobuta-1,3-diene and 1,4-bis(glutathion-\(\S_{\text{-yl}}\))-1,2,3,4- tetrachlorobuta-1,3-diene. Drug Metab. Dispos. 16, 701-706.
- Gietl, Y. S., and Anders, M. W. (1990) Biosynthesis and biliary excretion of S-conjugates of hexachlorobuta-1,3-diene in the perfused rat liver. <u>Drug Metab. Dispos.</u>, submitted.
- Vamvakas, S., Dekant, W., and Anders, M. W. (1989) Mutagenicity of benzyl <u>S</u>-haloalkyl and <u>S</u>-haloalkenyl sulfides in the Ames-test. <u>Biochem. Pharmacol.</u> 38, 935-939.
- Veltman, J. C., Dekant, W., Guengerich, F. P., and Anders, M. W. (1988) Cytotoxicity and bioactivation mechanism of benzyl 2-chloro-1,1,2-trifluoroethyl sulfide and benzyl 1,2,3,4,4-pentachlorobutadienyl sulfide. <u>Chem. Res. Toxicol.</u> 1, 35-40.

Reviews acknowledging AFOSR-86-0302:

- Anders, M. W. (1988) Glutathione-dependent toxicity: Biosynthesis and bioactivation of cytotoxic <u>S</u>-conjugates. <u>ISI Atlas of Science</u>: <u>Pharmacology</u> **2**, 99-104.
- Anders, M. W., Lash, L. H., Dekant, W., Elfarra, A. A., and Dohn, D. R. (1988) Biosynthesis and biotransformation of glutathione S-conjugates to toxic metabolites. CRC Crit. Rev. Toxicol. 18, 311-341.
- Anders, M. W., Vamvakas, S., and Dekant, W. (1990) Bioactivation of haloalkenes through glutathione conjugation. In <u>Glutathione S-transferases and Drug Resistance</u>, in press.
- Dekant, W., Lash, L. H., and Anders, M. W. (1988) Fate of glutathione conjugates and bioactivation of cysteine S-conjugates by cysteine conjugate β-lyase. In Glutathione Conjugation: Its Mechanism and Biological Significance. Sies, H.; Ketterer, B., eds., Academic Press, Orlando, pp. 415-447.
- Dekant, W., Vamvakas, S., and Anders, M. W. (1989) Bioactivation of nephrotoxic haloalkenes by glutathione conjugation: Formation of toxic and mutagenic intermediates by cysteine conjugate β-lyase. <u>Drug Metab. Rev.</u> 20, 43-83.
- Dekant, W., Vamvakas, S., and Anders, M. W. (1990) Bioactivation of hexachlorobutadiene by glutathione conjugation. <u>Drug Chem. Toxicol.</u>, in press.
- Monks, T. J., Anders, M. W., Dekant, W., Stevens, J. L., Lau, S. S., and van Bladeren, P. J. (1990) Glutathione conjugate mediated toxicities. <u>Toxicol. Appl. Pharmacol.</u>, in press.